## Evaluation of Metals in Soils used for Cultivation of *Rhamnus prinoides* (Gesho) at Bako Tibe District, Oromiya, Ethiopia

#### Ararso Nagari and Alemayehu Abebaw

Department of Chemistry, College of Natural and Computational Studies, Ambo University, Post Box No: 19, Ambo, Ethiopia. E-mail: ararsanago@gmail.com

#### Abstract

This study was carried out with the objective of determining the quantity of selected essential and non-essential metals; K, Na, Mg, Ca, Cu, Mn, Cr, Cd, Fe and Zn in the soil. Samples were collected from the low and medium altitudes of Bako (9°N latitude and 37.17°E). Wet acid-digestion using a mixture of aqua regia (1:3 HNO<sub>3</sub> to HCl) and H<sub>2</sub>O<sub>2</sub> (6:1 mL) was used. K and Na were analyzed using flame photometry, Ca and Mg were determined titrimetrically and the other metals with flame atomic absorption spectrometry (FAAS) after appropriate quality control measures were undertaken to verify and maintain the quality of the data generated. The results of the study showed that the average concentrations of K, Ca, Mg, Na, Cu, Mn, Fe, Zn, Cr and Cd were 2401.38, 8920, 633.19, 628.13, 125.79, 39.66, 38.51, 7.28 and 2.41 mg kg<sup>-1</sup> and below method detection limit, respectively in the low-altitude; and 3347.38, 10544, 1935.32, 758.13, 113.12, 25.41, 79.94, 50.95 and 3.52 mg kg<sup>-1</sup> and below method detection limit, respectively in the medium-altitude. The metals such as K, Ca, Mg, Na and Cu were generally found to be in higher concentrations than other metals in both samples.

Keywords: Bako Tibe, Flame atomic absorption spectrophotometer, metals, Soil.

#### Introduction

*Rhamnus prinoides* L'Herit, common name dogwood, Amharic name *Gesho*, family Rhamnaceae, is a wide spread plant species in East, Central and South African countries. The African dogwood, *R. prinoides* (Rhamnaceae) is a dense shrub or a tree that grows up to 6 m high. In Ethiopia, the leaves and stems of this plant are used to impart the characteristic bitter flavor to domestically brewed beverages known as *Tella* and *Tej* and it is estimated that well over 5 million people consume these beverages daily (Ararso and Alemayehu, 2013).

Soil is an essential medium for plant growth and for the breakdown and recycling of organic matter. Soil performs this function by supporting plants structurally and supplying elements essential for plant growth. An element is essential to a plant if its deficiency makes it impossible for the plant to complete the vegetative or reproductive stages of its life-cycle, the deficiency is specific to the

element in question and can be prevented or corrected by supplying this element and the element is directly involved in the nutrition of the plant. Essential elements are generally divided into major elements (macronutrients) and trace elements (micronutrients). Trace elements are as important as major elements but are required in lower quantities by plants. The trace elements are iron, manganese, boron. zinc, molybdenum, chlorine, cobalt, sodium and silicon. The maior elements considered essential for plant growth are carbon, hydrogen, nitrogen, phosphorus, oxygen, potassium, calcium, magnesium and sulphur (EPA, 2002).

Assessing pollutants in different components of the ecosystem has an important become task in preventing risk to natural life and public health (Pujar et al., 2012). Heavy metals contamination has received much attention with regard accumulation soils to in and aquatic contamination of environments (Pandey et al., 2011). In elemental this the study accumulations and distributions of selected major and trace metals (K, Na, Mg, Ca, Cr, Mn, Fe, Zn, Cu and Cd) in soil of low and medium altitudes of Bako Tibe district were investigated and compared with the metals concentration of stem and leaf of *Rhamnus prinoides* cultivated on the same soil in which all the samples were collected at the same time.

The objectives of the study were to: (i) determine the amounts of essential and non-essential metals in the soil, (ii) compare the amounts of essential and non-essential metals in the soil of low and medium altitude, and (iii) compare the amounts of essential and non-essential metals in the soil with the amounts in the stems and leaves of *Rhamnus prinoides*.

### **Materials and Methods**

# Description of the study area

Bako Tibe (Figure 1) is one of the districts in the Oromia Region of Ethiopia. Part of the West Shewa Zone, Bako Tibe is found at the Western part of the zone at about 250 km from the regional capital city and national capital, Addis Ababa, and 125 km away from the zonal capital, Ambo and bordered on the south and west by the East Welega Zone, on the north by Horo Gudru Welega Zone, and on the east by Ilu Gelan. The administrative center of this woreda is Bako.



Figure 1: Map of the Study Area

The woreda lies at an altitude of between 1500 and 2870 meters above sea level, of which 1500–1670 masl is considered low-altitude, 1670–2000 masl as medium-altitude and greater than 2000 masl as high-altitude.

# Apparatus, chemicals and reagents

Flame absorption atomic spectrophotometer (SL194, Double Elico) flame beam AAS, and photometer (CL 378, Elico) were used. HNO3 (65-68% Uni-Chem® Chemical reagents, India), extra pure hydrogen peroxide H<sub>2</sub>O<sub>2</sub> (30% Uni-Chem<sup>®</sup> Chemical reagents), aqua-regia (1:3, HNO3: HCl), HCl (35-38% LR, s.d.fine-cHEM LimTEd, MUMBAI) and sucrose, C12H22O11 (HIMEDIA® A.R.), stock standard solutions (1000 ppm) prepared in 20 mL 1:1HNO<sub>3</sub> for Cu metal diluted to 1 L, 17 mL 1:1 HCl for Zn and Fe diluted to 1 L and distilled water were used in this study.

#### **Cleaning of apparatus**

All the glassware and materials used during sample collection and experimental work were cleaned before and after use with tap water and detergent, rinsed with distilled water, soaked with 10% (v/v) nitric acid, re-rinsed with distilled water and subjected to total air dry to ensure decontamination.

#### Stock and working intermediate metal standard preparation

Stock standard solutions of each of the metals of interest were prepared from high purity metals and reagent grade salts using distilled water, nitric acid and hydrochloric acid based on the individual analysis specific instruction of the instruments. The stock solutions were prepared at concentrations of 1000 mg mL<sup>-1</sup> of the metal.

The working intermediate metal standard solutions (100  $\mu$ g mL<sup>-1</sup> each) were prepared by diluting 10 mL of 1000  $\mu$ g mL<sup>-1</sup> individual metal stock standard solutions with distilled water to a 100 mL volume.

# Calibration metal standard solutions

A blank and five calibration standard solutions of different concentration levels were prepared for each metal working the respective from intermediate standard solutions (100 µg mL<sup>-1</sup>). The metal concentrations include 1.5, 2.5, 3.5, 4.5, 5.5 µg mL<sup>-1</sup> for Cr; 0.25, 0.5, 0.75, 1.0 1.25 µg mL<sup>-1</sup> for Cd; 1.0, 2.0, 3.0, 4.0, 5.0 µg mL<sup>-1</sup> for Cu; 1.0, 1.75, 2.5, 3.25, 4.0 µg mL<sup>-1</sup> for Mn; and 0.5, 1.0, 2.0, 4.0, 8.0 µg mL<sup>-1</sup> for Fe and Zn. These calibration standard concentrations within the were working linear range of the instrument used for analysis. Beginning with the blank and working toward the highest standard, the solutions were aspirated and the readings were recorded.

### Spiking metal standard mixture solution

Standard mixture solution containing all the metals of interest at varying concentrations (K and Na = 80, Mn = 40, Cu and Cr = 50, Fe and Zn = 60, Cd = 10  $\mu$ g mL<sup>-1</sup>) was prepared from stock standard solutions in a 50 mL volumetric flask. The volume of spike stock solution that should be added to the sample was calculated.

### Sample collection and processing

Purposeful sampling technique was used to collect soil samples from the low and medium altitude areas of the studv area. Soil samples were specifically collected from around the plant and composited (McDonald et al., 2004), in such a way as to be representative of the surface soil layers from a 20 cm depth. Nondecomposed organic materials were removed prior to sampling (El-Ghawi et al., 2005). The samples collected in January, 2013. The sub-samples were transported in plastic bags to Ambo University chemistry laboratory for further treatment and analysis. The samples were air dried to constant weight for two weeks, grinded using mortar and pestle and sieved through a 1.5 mm stainless steel sieve. The 0.5 g from the sieved sample were digested and analyzed.

## Determination of moisture correction factor

The samples were weighed with analytical electronic balance to record the initial weight. It was re-weighed and re-dried until it gives constant mass after oven drying at 105 °C for 12 hrs and the moisture content was

determined from the differences and the corresponding moisture correction factor for analytical results or the multiplication factor for the amount of sample to be weighed for analysis was calculated (FAO, 2008).

# Determination of soil pH in water

Soil pH was normally measured in soil-water slurry. The determination of the soil pH is a three-step procedure: first, the soil-water slurry in the ratio 1:5 was prepared; and then, the pH meter was calibrated with a buffer solution of pH 4, 7 and 9; and finally, the electrodes were placed into the prepared slurry and the pH was read and recorded (FAO, 2008; Benton, 2001).

# Optimization of digestion procedure for soil samples

The measurement of metal concentration in soil was generally by a combination of digestion procedure for dissolution of elements and subsequent measurement the of dissolved elements as described by Gaudino et al., (2007). The conventional aqua-regia digestion procedure as described by Chen and Ma (2001). For digesting soil samples was adopted.

The mixture of aqua-regia (HNO<sub>3</sub> + 3HCl) and H<sub>2</sub>O<sub>2</sub> was selected as digestion reagent for the soil samples in this stuidy. The digestion procedures were optimized using different proportions of aqua-regia and H<sub>2</sub>O<sub>2</sub> mixtures by varying the

digestion time and temperature. The optimum condition for soil samples digestion was found to be a mixture of 6mL aqua-regia (3:1 ratio of HCl to HNO<sub>3</sub>) and 1mL H<sub>2</sub>O<sub>2</sub>, with a digestion temperature of 220°C and digestion time of 2 hrs.

#### Quality assurance/control Calibration and continuing calibration standard

The FAAS was calibrated using a blank and five series of working standard solutions of each metal. The continuing calibration standard of metal standard solution of mid-point calibration concentration was run to test the accuracy of the analytical instrument calibration with each batch samples as described by Csuros and Csuros, (2002) and the deviation from the original value was found to be within ±5%.

The performance of flame photometer for K and Na was checked with the freshly prepared standard solutions of two respective the metals at concentrations of 10, 30, 50, 70, 90 ppm for K and at 1, 5, 10, 20 ppm for Na. The corresponding recovered/ measured values 9.57±1.69, 31.03±1.99, 50.07±0.47, 69.63±0.71 and 89.27±1.27 for K and 0.98±0.10, 5.03±0.15, 10.13±0.31, 15.33±0.76 and 20.67±0.86 for Na were obtained indicating a good performance of the instrument.

# Analysis of matrix spike and matrix spike duplicate

Both matrix spike (MS) and matrix spike duplicate (MSD) were prepared

for each soil sample by spiking aliquots of 0.5 g of soil with 2.5 mL standards mixture solution equivalent to a concentrations of 4.0 µg mL<sup>-1</sup> for K and Na, 2.5 µg mL<sup>-1</sup> for Cu and Cr, 0.5 µg mL<sup>-1</sup> for Cd and 3.0 µg mL<sup>-1</sup> for They were all carried Fe and Zn. through the same digestion and analysis steps as an un-spiked sample. The mean recovery values of MS and MSD were used to control overall sample matrix-dependent accuracy of the analytical method. Their relative percent differences (RPD) were calculated and expressed as percent to continuously assess the corresponding method precision (Csuros and Csuros, 2002; Miller and Miller, 2010).

#### **Precision and accuracy**

The laboratory precision of the results was assessed by the analysis of laboratory control samples (LCSs, n=3), laboratory control sample duplicates (LCSD), MS (n=3) and MSD. Precision was expressed as the RPD and relative standard deviation (RSD) of replicate results.

The accuracy of the optimized procedure was evaluated by spiking experiments. This included analysis of the MS (n=3), MSD, LCSs (n=3), LCSD and method blanks. Accuracy was expressed as the percent recovery of an analyte that has been spiked to the samples in a known concentration before digestion and subsequent analysis.

### Method blank, reagent blank and limit of detection and quantification

The method blank was prepared from sucrose as matrix since there was no clear soil that can serve as the matrix for the soil samples. The blank which was prepared from the sucrose and all reagents was used for the digestion through and taken the entire measurement procedure to detect contamination from reagents, sample handling, and the entire measurement process. Method detection limit (MDL) and limit of quantification for each metal of interest were estimated by digesting and analyzing seven replicates of the method blanks with the optimized procedure for the soil samples. While the instrument was run with reagent blanks/calibration blanks in which no sample was used to determine the background signal and establish the baseline of the instrument.

#### Analysis of laboratory control samples

Triplicates of LCSs prepared from sucrose spiked with the target analyte of known concentration the same to sample spike were analyzed along with each batch of the soil samples. LCS recoveries and the RSD values of the LCS recoveries values for each metal of interest were calculated and used to control overall analytical method accuracy and precision, respectively which were independent

of sample matrix effects (Csuros and Csuros, 2002; Miller and Miller, 2010).

## Analysis of metal levels in soil samples

Five replicate analyses were carried out on each sample for the determinations of Na and K with flame photometry and Cu, Mn, Cr, Cd, Fe and Zn with FAAS; and the concentrations of Ca and Mg were determined by complexometric EDTA titration following the FAO procedure (FAO, 2008).

#### **Statistical analysis**

The variation in sample mean of the analyte was tested by using T-test of SPSS Version20 Software to test whether the source for variation was from experimental procedure or heterogeneity among the samples. Excel 2007 was used for other calculations.

### **Results and Discussion**

## Quality assurance/control Calibration

Calibration curves for the various concentration ranges showed good correlation coefficients ranged between 0.996 and 0.999 (Table 3), which were all within the required limit (0.995) for trace element analysis (US EPA, 2007). This showed that there was good correlation (relationship) between concentration and absorbance indicating good calibration of the instrument.

## Matrix spike and matrix spike duplicate

highest The average recovery obtained was 100.75% (Na) and the lowest 90.5% (Zn) (Table 1) where all were in the range 80-120%, which is recommended for matrix spike recovery of major element analyses by FAAS and Flame spectrophotometer (US EPA, 2008). This indicated that the analytical process has provided matrix-dependant required the accuracy. Therefore, the method was well applicable for all the matrices and all the results obtained were trustable

RPD values of MS and MSD obtained ranged between 1.631 (Zn) and 13.40 (Na) which were in the acceptance limit ( $\leq$ 15%) (Csuros and Csuros, 2002). This showed that the overall analytical process was less affected by matrix-specific variability and the data generated was in the acceptable quality range.

| Metals | LCS (%R, n=3) | LCS (RSD) | MS & MSD (%R, n=2) | MS & MSD (RPD) |
|--------|---------------|-----------|--------------------|----------------|
| К      | 91.667±3.82   | 4.17      | 92.375 ±3.71       | 5.68           |
| Na     | 87.5±9.01     | 10.30     | 100.75 ±9.55       | 13.40          |
| Cu     | 93.21±7.16    | 7.68      | 91.10 ±0.27        | 10.69          |
| Mn     | 94.57±4.44    | 4.69      | 98.2 ±3.96         | 3.10           |
| Cr     | 98.56±2.86    | 2.90      | 98.77 ±2.61        | 4.55           |
| Cd     | 96.58±4.08    | 4.23      | 98.4 ±4.81         | 2.78           |
| Fe     | 97.56±1.73    | 1.77      | 97.73 ±0.43        | 2.59           |
| Zn     | 96.78±1.97    | 2.04      | 90.5 ±0.17         | 1.63           |

Table 1: Mean %R and RPD values of metals from MS and MSD; and %R and RSD values of LCSs analyzed along with real samples for method validation

LCS= laboratory control samples, %R= percent recovery, RSD= relative standard deviation, MS= matrix spike, MSD= matrix spike duplicate, RPD= relative percent difference

#### **Method precision and**

#### accuracy

The calculated RSD of recovery values were under the required limit (Table 1). This is within  $\leq 15\%$  as reported by Csuros and Csuros (2002). This indicated that the analytical method, which digestion covers and instrumental measurement has acceptable repeatability and precision.

The percent recoveries of the sample spiked ranged between the highest 99.25% (Na) and lowest 93.08% (K) (Table 2). All were found to be within the acceptable range (80–120%) for metal analysis (US EPA, 2008). This showed that the analytical method provided results in the required level of accuracy.

Table 2: Recovery and precision test for the optimized procedure from soil sample spikes (n = 3)

| Metals | Conc. in sample<br>(µg mL <sup>-1</sup> ) | Amount spiked<br>(µg mL <sup>-1</sup> ) | Conc. in spiked sample (µg mL-1) | Recovery (%) | Precision<br>(%, RSD) |
|--------|---|---|----------------------------------|--------------|-----------------------|
| K      | 24.03±0.84                                | 4.0                                     | 27.76±0.91                       | 93.08±2.90   | 3.11                  |
| Na     | 6.43±0.21                                 | 4.0                                     | 10.40±0.44                       | 99.25±7.23   | 7.29                  |
| Cu     | 37.71±2.14                                | 2.5                                     | 39.60±2.13                       | 96.65±3.75   | 5.38                  |
| Mn     | 26.30±0.87                                | 2.0                                     | 28.27±0.84                       | 98.38±2.82   | 2.96                  |
| Cr     | 4.19 ±0.33                                | 2.5                                     | 6.56 ±0.39                       | 95.10 ±2.57  | 5.90                  |
| Cd     | 0.61±0.05                                 | 0.5                                     | 1.09±0.02                        | 96.33±6.11   | 1.91                  |
| Fe     | 21.58±1.47                                | 3.0                                     | 24.47±1.48                       | 96.18±1.87   | 6.05                  |
| Zn     | 11±1.52                                   | 3.0                                     | 13.92±1.38                       | 97.44±4.81   | 9.89                  |

RSD= Relative standard deviation

#### Instrument detection, method detection and quantification limits

The reagent blank readings were below the method detection limit indicating that no instrumental variations during the measurements. The method blank analysis results revealed that there were no readings above the method detection limits of the metals. Hence, the analytical method was free from overall laboratory contamination.

The instrument detection limits for both FAAS and FES ranged between 0.06 and 0.72 mg kg-1, indicating good sensitivity measuring of the instrument for analyses. The MDL values lied in the range 0.76 (Cd)-3.9 (K) mg kg<sup>-1</sup> and method quantification limit values 1.68 (Cd)-6.91 (K) mg kg-1 (Table 3). All the values were found to be in the required limit (Temminghoff and Houba, 2004). This indicated that the method was well applied for the determination of the essential and non essential metals.

Table 3: Instrument detection limit (IDL), method detection limit (MDL), method quantification limit (MQL) and correlation coefficients of calibration curves for metals determined in the soil samples

| Metals | Correlation coefficient of<br>calibration curves (r <sup>2</sup> ) | Calibration equation | IDL (mg kg <sup>-1</sup> ) | MDL<br>(mg kg <sup>-1</sup> ) | MQL (mg kg <sup>-1</sup> ) |
|--------|--|----------------------|----------------------------|-------------------------------|----------------------------|
| К      | -  | -                    | 0.72                       | 3.90                          | 6.91                       |
| Na     | -  | -                    | 0.40                       | 2.66                          | 4.50                       |
| Cu     | 0.999  | Y= 0.033X+0.003      | 0.62                       | 3.12                          | 5.52                       |
| Mn     | 0.996  | Y= 0.008X+0.003      | 0.15                       | 1.89                          | 5.50                       |
| Cr     | 0.999  | Y= 0.013X+0.002      | 0.13                       | 0.77                          | 1.99                       |
| Cd     | 0.999  | Y= 0.082X+0.002      | 0.06                       | 0.76                          | 1.68                       |
| Fe     | 0.998  | Y= 0.015X+0.005      | 0.59                       | 3.36                          | 4.59                       |
| Zn     | 0.997  | Y= 0.049X+0.020      | 0.44                       | 3.30                          | 6.04                       |

#### Laboratory control sample

The percent recovery values of LCS lied in the range 87.5 (Na)–98.56% (Cr) and their relative standard deviations 1.77 (Fe)–10.30 (Na) (Table 4). And all the values were found under standard control limits 80–120% for LCS

recovery, and ≤15% for RSD (Csuros and Csuros, 2002). This showed that the method used for the study has provided the required level of accuracy and precision throughout the analytical process.

| Metals | Amount spiked<br>(mg L <sup>-1</sup> ) | Conc. in spiked sample (mg L-1) | Accuracy<br>(Recovery, %) | Precision<br>(RSD, %) |  |  |
|--------|--|---------------------------------|---------------------------|-----------------------|--|--|
| К      | 4.0                                    | 3.67 ±0.15                      | 91.67 ±3.82               | 4.17                  |  |  |
| Na     | 4.0                                    | 3.5 ±0.36                       | 87.5 ±9.01                | 10.30                 |  |  |
| Cu     | 2.5                                    | 1.9 ±0.13                       | 93.21 ±7.16               | 7.68                  |  |  |
| Mn     | 2.0                                    | 1.89 ±0.09                      | 94.57 ±4.44               | 4.69                  |  |  |
| Cr     | 2.5                                    | 2.46 ±0.07                      | 98.56 ±2.86               | 2.9                   |  |  |
| Cd     | 0.5                                    | 0.48 ±0.02                      | 96.58 ±4.08               | 4.23                  |  |  |
| Fe     | 3.0                                    | 2.93 ±0.05                      | 97.56 ±1.73               | 1.77                  |  |  |
| Zn     | 3.0                                    | 2.9 ±0.06                       | 96.78 ±1.97               | 2.04                  |  |  |

Table 4: Percent recoveries and RSD values of laboratory control soil samples analyzed along with real samples (n = 3)

# Soil pH and metals in the soil samples

The pH of the low altitude soil was  $6.82 \pm 0.22$  and that of the medium altitude was  $7.21\pm0.26$ . The low altitude soil was slightly acidic while the medium altitude soil can be described as being moderately alkaline.(FAO, 2008).

The moisture content of the two soil samples were 2.55 and 1.26% with moisture correction factor (mcf) of 1.03 and 1.01 respectively for both medium and low altitude areas. These correction factors were used for the calculation of the metal concentrations.

The total concentrations of the metals in the soil samples are presented in Table 5. The mean concentrations of all metals were higher in both the medium altitude and lower altitude soil samples except chromium. However, in the lowland altitude all metal concentrations were found to be lower when compared to that of the medium altitude, with the exception of manganese which recorded a higher value (39.66 mg kg-1 ) in the lower altitude. T-test showed significant differences (P < 0.05, at 95% CL) among the analyzed soil samples for the mean concentrations of all metals except for Cu. The concentrations of all metals except Cr in the analyzed soil samples were under the EPA maximum permissible limit of typical concentration in soil dry matter 1,000-30,000 mg kg-1 for K, 5,000-30,000 mg kg<sup>-1</sup> for Ca, 1,000-15,000 mg kg<sup>-1</sup> for Mg, 10,000–50,000 mg kg<sup>-1</sup> for Fe, 20-30 mg kg<sup>-1</sup> for Mn, 2-100 mg kg-1 for Cu, 10-200 mg kg-1 for Zn, 0.1–1 mg kg<sup>-1</sup> for Cd (EPA, 2002) and the minimum reporting limit 1 mg kg-1 for Cr in the soil (US EPA, 2012).

The distribution of the selected essential and non-essential metals in each soil samples were in the order of: Ca > K > Mg > Na > Cu > Mn > Fe > Zn > Cr > Cd (BMDL) in the low altitude soil samples and Ca > K > Mg > Na > Cu > Fe > Zn > Mn > Cr > Cd (BMDL) in the medium altitude soil samples.

[74]

Table 5: Average concentration (mg kg<sup>-1</sup> air dry weight) of metals in low and medium altitude soil samples

|                                     | Average concentrations (mg kg-1 air dry weight) |               |               |                 |               |            |              |      |              |             |
|-------------------------------------|---|---------------|---------------|-----------------|---------------|------------|--------------|------|--------------|-------------|
| Sample type                         | К   | Na            | Ca            | Mg              | Cu            | Cr         | Mn           | Cd   | Fe           | Zn          |
| Low altitude soil                   | 2401.38 ±95.03                                  | 628.13 ±27.39 | 8920 ±126.49  | 633.19 ±60.96   | 125.79 ±4.98  | 2.41 ±0.24 | 39.66 ±0.99  | BMDL | 38.51 ±10.11 | 7.28 ±2.95  |
| Medium altitude soil                | 3347.38 ±263.82                                 | 758.13 ±45.83 | 10544 ±121.98 | 1935.32 ±105.89 | 113.12 ±14.92 | 3.52 ±0.57 | 25.407 ±0.19 | BMDL | 79.94 ±9.91  | 50.95 ±9.34 |
| BMDL= below method detection limit, |   |               |               |                 |               |            |              |      |              |             |

The analyzed selected metal levels in low altitude soil sample when compared with that of stem and leaf of the Rhamnus prinoides showed that significantly thev were varied (Ararsa and Alemayehu, 2013). The concentration of K was highest in leaf and lowest in soil. Na, Mg, Cu, Fe and Cr levels were highest in soil, lowest in leaf; Ca and Mn were higher in soil, lower in the stem; Zn levels were found to be lower in the leaf and higher in the stem samples; however, Cadmium levels were below method detection limits in all samples(Table 6).

In a related study of the distributions of essential and non-essential metal levels in the medium-altitude soil of Bako Tibe , the concentrations of metals in the stem and leaf of *Rhamnus prinoides* were also found to vary significantly (Ararso and Alemayehu, 2013). The concentration of K and Ca were higher in leaf and lowest in soil. Na, Fe, Zn and Cu levels were higher in soil and lower in leaf. Ca was found to be lower in stem when compared to the leaf and that of the concentration in soil. Moreover, the concentrations of Mg, Mn and Cr were higher in soil and lower in the stem. Cadmium levels were below method detection limits in all the samples (soil and plant) (Table 6).

Thus, from this study, most of the accumulation of metals in the stem and leaf of *Rhamnus prinoides* could be attributed to their presence in the soils on which the plant was being cultivated in addition to other factors such as soil pH and the use of manure. Generally, concentrations were found to be higher in the medium altitude compared to the lower altitude areas. This could be attributed to higher rainfall, more vegetation and level of organic matter decomposition in addition to the soil pH.

Table 6: Comparison of average concentration (mean ± SD, n = 5, mg kg<sup>-1</sup> oven dry weight) of metals in stem and leaf of *R. prinoides* and soil samples

| Metals - | Leaf (mg kg <sup>-1</sup> oven dry weight) |                  | Stem (mg kg <sup>-1</sup> | oven dry weight) | Soil (mg kg <sup>-1</sup> | Soil (mg kg <sup>-1</sup> oven dry weight) |  |
|----------|--|------------------|---------------------------|------------------|---------------------------|--|--|
|          | Low altitude                               | Medium altitude  | Low altitude              | Medium altitude  | Low altitude              | Medium altitude                            |  |
| К        | 12927.3 ± 346.92                           | 11651.3 ± 333.44 | 10239.54 ± 386.23         | 8855.54 ± 269.94 | 2401.38 ± 95.03           | 3347.38 ± 263.82                           |  |
| Na       | 226.21±19.24                               | 320.21 ±10.37    | 308.66 ±7.91              | 413.66±7.91      | 628.13 ± 27.39            | 758.13 ±45.83                              |  |
| Са       | 7424 ±159                                  | 11120 ±126.49    | 6144 ± 191                | 7416 ± 153.88    | 8920 ±126.49              | 10544 ±121.98                              |  |
| Mg       | 352.34 ±13.14                              | 1526.81 ±122.98  | 413.62 ± 12.64            | 1210.21 ±103.08  | 633.19 ± 60.96            | 1935.32 ±105.89                            |  |
| Cu       | 29.1 ± 3.46                                | 36.98 ±6.43      | 46.34 ± 4.75              | 49.91 ± 5.83     | 125.79 ± 4.98             | 113.12 ±14.92                              |  |
| Mn       | 13.11 ± 0.28                               | 7.36 ±1.98       | 9.41 ± 0.26               | $3.36 \pm 0.2$   | 39.66 ±1.0                | 25.41 ± 0.19                               |  |
| Cr       | 1.71 ± 0.34                                | 1.71 ±0.19       | 2.17 ± 0.3                | 2.37 ± 0.24      | 2.41 ± 0.24               | 3.52 ± 0.57                                |  |
| Cd       | BMDL                                       | BMDL             | BMDL                      | BMDL             | BMDL                      | BMDL                                       |  |
| Fe       | 8.58 ±1.23                                 | 10.71 ±3.98      | 9.39 ± 3.37               | 10.73 ± 0.1      | 38.51 ±10.11              | 79.94 ± 9.91                               |  |
| Zn       | 3.48 ±1.54                                 | 7.91 ±4.99       | 16.33 ± 3.04              | 18.36 ± 6.12     | 7.28 ±2.95                | 50.95 ± 9.34                               |  |

BMDL = below method detection limit

## Conclusion

A wet acid-digestion method using the mixture of aqua-regia (HCl and HNO<sub>3</sub>) and H<sub>2</sub>O<sub>2</sub> for preparation of samples were optimized, soil validated and used. The determinations of essential and non-essential metal levels (K, Na, Ca, Mg, Cr, Cu, Zn, Fe, Mn and Cd) in the samples were by flame photometer, flame atomic absorption spectrophotometer and complexometric titration with EDTA. The quality of the analytical data generated and general laboratory continuously performance were monitored and verified through the quality control parameters including CCS, LCSs, MS, MSD, method blank solvent/reagent laboratory and blanks. The laboratory procedures were all within acceptable limits/ guidelines.

Elements such as K, Ca, Mg, Na and Cu were generally found in higher concentrations than the other metals of interest in both samples. There was variations in metal concentrations in the soil which may be ascribed to: the physical and chemical nature of the soil (cation exchange capacity, organic and soil matter content рН); atmospheric deposition of the metals (which may be influenced bv innumerable environmental factors such as temperature, moisture and pH) (Kalra, 1998).

## Acknowledgement

The authors express their deepest appreciation and thanks to Ambo University for providing the fund and laboratory facility.

### References

- Ararso., N. and Alemayehu., A. 2013. Determination of Selected Essential and Non-essential Metals in the Stems and Leaves of *Rhamnus prinoides* (Gesho). *STAR Journal*, 2(4): 20-26.
- Benton., J. J. 2001. Laboratory Guide for Conducting Soil Tests and Plant Analysis. CRC Press LLC, Boca Raton, USA, 382Pp.
- Chen., M, and Ma L. Q. 2001. Comparison of three aqua-regia digestion methods for Florida Soils. Soil Science. Society of America Journal, 65: 491–499.
- Csuros., M, and Csuros., C. 2002. Environmental sampling and analysis for metals. CRC Press, USA.
- El-Ghawi, U. M., Bejey, M. M., Al-Fakhri., S. M., Al-Sadeq., A. A. & Doubali., K. K. 2005. Analysis of Libyan arable soils by means of thermal and epithermal NAA. Arabian Journal for Science and Engineering, 30 (1A) : 147–153.
- Environmental Protection Agency (EPA) 2002. Setting environmental quality objectives for soil: Developing a soil protection strategy for Ireland. A discussion document by Ms. Jane Brogan, Dr Matt Crowe and Mr. Gerry Carty. Johnstown Castle Estate, County Wexford, Ireland.
- Food and Agricultural Organization (FAO) 2008. Guide to Laboratory Establishment for Plant Nutrient

Analysis Fertilizer and Plant Nutrition Bulletin No.19.

- Gaudino., S., Galas., C., Bell, M., Barbizzi., S., Zorzi, P., Jacimovic., R, Jeran, Z., Pati, A., & Sansone., U. 2007. The Role of Different Soil Sample Digestion Methods on Trace Elements Analysis: a Comparison of ICP-MS and INAA Measurement Results. J. Qual. Comp. Reliability Chem. Meas. 12: 84–93.
- Kalra., Y. P. 1998. Handbook of Reference Methods for Plant Analysis. Taylor & Francis Group, Boca Raton, USA. 291Pp
- McDonald., L, Slagle., A., Skousen., J., Bhumbla., D. & Sencindiver., J. 2004. Trace Element Concentrations of three soils in central Appalachia. Soil Sur. Hori. Fall 45(3): 73–85.
- Miller, N. J. and Miller., C. J. 2010. Statistics and Chemometrics for Analytical Chemistry, 6<sup>th</sup> ed. Pearson Education Limited, UK. 297Pp
- Pandey, R., Meena., O., Garg, A. & Singh., M. 2011. Determination of toxic trace metals (Pb, Cd, Ni, and Zn) in Soil by

Polarographic Method. Int. J. Chem. Tech. Res, 3 (2): 599–604.

- Pujar., K. G., Hiremath, S. C, Pujar., A. S., Pujeri, U. S. & Yadawe., M. S. 2012. Analysis of Physico-chemical and Heavy Metal Concentration in Soils of Bijapur Taluka, Karnataka. Sci. Revs. Chem. Comm. 2(1): 76–79.
- Temminghoff., E. J. and Houba., V. J. 2004. Plant Analysis Procedures Second Edition. Kluwer Academic Publishers, Netherlands. 187Pp
- United State Environmental Protection Agency (USEPA) 2007. Solutions to Analytical Chemistry Problems with Clean Water Act Methods. EPA 821-R-07-002, Washington, DC.
- United State Environmental Protection Agency (USEPA) 2008. National Functional Guidelines for Superfund Organic Methods Data Review. USEPA-540-R08-01, Washington, DC.
- United State Environmental Protection Agency (USEPA) 2012. Laboratory operations and quality assurance manual. Analytical support branch, Athens, Georgia 30605–2700.